Seeds of future past: climate change and the thermal memory of plant reproductive traits

Eduardo Fernández-Pascual^{1,2*}, Efisio Mattana³ and Hugh W. Pritchard¹

¹Comparative Plant and Fungal Biology, Royal Botanic Gardens, Kew; Wellcome Trust Millennium Building, Wakehurst Place, Ardingly, West Sussex RH17 6TN, UK ; ²Departamento de Biología de Organismos y Sistemas, Universidad de Oviedo, C/ Catedrático Rodrigo Uría, 33006 Oviedo/Uviéu, Spain; ³Natural Capital and Plant Health, Royal Botanic Gardens, Kew; Wellcome Trust Millennium Building, Wakehurst Place, Ardingly, West Sussex RH17 6TN, UK

*Author for correspondence (E-mail: eduardofp.indurot@uniovi.es; Tel.: +34985104787).

This is the peer reviewed version of the following article: Fernández-Pascual E, Mattana E, Pritchard HW (2019) Seeds of future past: climate change and the thermal memory of plant reproductive traits. Biological Reviews 94, 439–456, which has been published in final form at <u>http://dx.doi.org/10.1111/brv.12461</u>. This article may be used for non-commercial purposes in accordance with Wiley Terms and Conditions for Use of Self-Archived Versions.

ABSTRACT

Plant persistence and migration in face of climate change depend on successful reproduction by seed, a central aspect of plant life that drives population dynamics, community assembly and species distributions. Plant reproduction by seed is a chain of physiological processes, the rates of which are a function of temperature, and can be modelled using thermal time models. Importantly, while seed reproduction responds to its instantaneous thermal environment, there is also evidence of phenotypic plasticity in response to the thermal history experienced by the plant's recent ancestors, by the reproducing plant since seedling establishment, and by its seeds both before and after their release. This phenotypic plasticity enables a thermal memory of plant reproduction, which allows individuals to acclimatise to their surroundings. This review synthesises current knowledge on the thermal memory of plant reproduction by seed, and highlights its importance for modelling approaches based on physiological thermal time. We performed a comprehensive search in the Web of Science and analysed 533 relevant articles, of which 81 provided material for a meta-analysis of thermal memory in reproductive functional traits based on the effect size Zr. The articles encompassed the topics of seed development, seed yield (mass and number), seed dormancy (physiological, morphological and physical), germination, and seedling establishment. The results of the meta-analysis provide evidence for a thermal memory of seed yield, physiological dormancy and germination. Seed mass and physiological dormancy appear to be the central hubs of this memory. We argue for integrating thermal memory into a predictive framework based on physiological time modelling. This will provide a quantitative assessment of plant reproduction, a complex system that integrates past and present thermal inputs to achieve successful reproduction in changing environments. The effects of a warming environment on plant reproduction cannot be reduced to a qualitative interpretation of absolute positives and negatives. Rather, these effects need to be understood in terms of changing rates and thresholds for the physiological process that underlie reproduction by seed.

Key words: climate warming, dormancy, functional traits, germination, global change, phenotypic plasticity, seed mass and number, seedling establishment, thermal time.

CONTENTS

ABSTRACT	1
I. INTRODUCTION	
(1) Climate warming and plant reproduction	
(2) Thermal time and plant reproduction	
II. META-ANALYSIS METHODOLOGY	
(1) Literature search	

(2) Data extraction and calculation of effect sizes	,
(3) Phylogenetic correlation matrices	
(4) Multi-variate meta-analysis models)
(5) Heterogeneity in effect sizes	j
(6) Publication bias	;
(7) Sensitivity analysis	;
III. META-ANALYSIS OF THE THERMAL MEMORY OF PLANT REPRODUCTIVE TRAITS 6	;
(1) General meta-analysis diagnostics	j
(2) Reproductive phenology	,
(3) Seed yield: mass and number	,
(4) Seed dispersal	;
(5) Morphological dormancy and embryo growth	;
(6) Physical dormancy and coat permeability	;
(7) Primary physiological seed dormancy and germination traits	;
(8) Post-dispersal seed dormancy cycles)
(9) Seedling growth)
IV. SYNTHESIS)
(1) The first hub of thermal memory: seed yield)
(2) The second hub of thermal memory: seed dormancy)
(3) Implications for modelling: an example)
(4) Future priorities	
V. CONCLUSIONS	-
VI. ACKNOWLEDGEMENTS 11	
VII. REFERENCES	

I. INTRODUCTION

In a climate change context, the ability of a plant species to persist in its current habitat or to migrate to new sites will depend on successful reproduction by seed. Seed traits affect all levels of plant ecology from population dynamics (Huang et al., 2016) to community assembly (Larson & Funk, 2016) and species distributions (Bykova et al., 2012). Seeds are also the basis for human activities such as agriculture, ex situ conservation (Li & Pritchard, 2009) and ecological restoration (Merritt & Dixon, 2011). Recently, Walck et al. (2011) highlighted how climate change is altering the major postdispersal seed-germination cues: temperature and water availability. However, seed responses to these cues are not fixed species traits. As Cochrane et al. (2015) showed, there is ample intraspecific variability in seed traits, which can provide the raw material for acclimatisation and/or adaptation to new climates. Intraspecific variability can be the result of either genetic differences (Fernández-Pascual et al., 2013) or phenotypic plasticity (Fenner, 1991), but the latter provides the fastest way for plants to adjust to their environment (Nicotra et al., 2010). Importantly, phenotypic plasticity allows plants to take into account their thermal history, integrating both the pre- and the post-dispersal environments into cross-generational plasticity (Auge et al., 2017). We define thermal history as the past thermal environments experienced by the seed and its recent ancestors (generally the parental and grandparental generations), as opposed to the current thermal environment reviewed by Walck et al. (2011). Phenotypic plasticity in response to thermal history amounts to a 'thermal memory' of plant reproductive traits. Remembering their thermal history would inform seeds to germinate in the most convenient conditions for seedling establishment and allow individuals to acclimatise to new surroundings.

This review has the double objective of promoting a physiological thermal time framework to study plant reproduction, and highlighting the importance of thermal memory in such a framework. As ectotherm life forms, most plant physiological processes are a function of environmental temperature, and suitable for quantification using a thermal time approach (Garcia-Huidobro, Monteith, & Squire, 1982). Each process has an optimal temperature at which its rate is maximal. Below and above this optimum, the rate progressively decreases until the temperature reaches base and ceiling thresholds beyond which the process stops. The optimal, base and ceiling temperatures are the cardinal temperatures of this process. For the process to be completed, a certain number of thermal units must accumulate, but this accumulation only happens within the limits of the cardinal temperatures. As we will see in this introduction, these thermal time concepts apply to the successive steps of plant reproduction by seed,

from seed production to seedling emergence. Through these concepts, plant reproduction becomes measurable by a set of thermal time traits: the cardinal temperatures and the required thermal time. This approach contributes to a predictive framework for plant reproduction, one that relates directly to the environmental thermal inputs via process-based models (Donohue et al., 2015). But if we want to use this framework to predict plant reproduction under changing climates, it is essential to acknowledge that thermal time traits are not fixed species traits but have a thermal memory. To support this critical point, this review synthetises the available evidence on the thermal memory of plant reproductive traits by means of a meta-analysis. We begin by providing a brief state-of-the-art regarding plant reproduction under climate warming; followed by an exploration of thermal time models of plant reproduction. We then perform a meta-analysis of the thermal memory of plant reproductive traits. Finally, we integrate current knowledge towards a predictive framework of plant reproduction, and highlight future avenues for research.

(1) Climate warming and plant reproduction

Evidence of the effect of climate change on plant reproduction is accumulating rapidly. Warming will generally advance plant reproduction in a wide variety of habitats (Wookey et al., 1993; De Frenne et al., 2011), but delays are also possible as a result of regional specificities (Del Cacho, Peñuelas & Lloret, 2013b; Bjorkman et al., 2015). These changes in reproductive phenology will have a cascading effect on the phenology and thermal history of subsequent life stages. Seed yield (i.e. the reproductive output of plants) will increase or decrease depending on whether the current limiting factor for a species is temperature (Richardson et al., 2005) or drought (Mutke, Gordo & Gil, 2005). Once seed production is affected, dispersal limitations will determine the future distributions of plant species (Engler et al., 2009), and dispersal distances themselves will be modified by changing wind patterns (Kuparinen et al., 2009). Climate warming will likewise disrupt the natural cycles of seed dormancy (Ooi, Auld & Denham, 2009), which is in essence a form of dispersal in time. The germination of non-dormant seeds will also be affected, with faster rates expected in warm-cued seeds from cold regions (Milbau et al., 2009) and delayed emergence in cold-cued seeds from dry regions (Cochrane, Daws & Hay, 2011). Above all, we must stress that a higher germination percentage – and an advanced or delayed emergence – are not positive or negative per se: the essential question is whether emergence will stay matched with the favourable establishment season. Moderate warming, by itself, tends to favour seedling establishment (De Frenne et al., 2012; Cheesman & Winter, 2013; Stevens et al., 2014; Mondoni *et al.*, 2015). But a seedling that emerges before its time can find itself dealing with frost-thaw events (Huelber, Bardy & Dulinger, 2011), drought (Del Cacho *et al.*, 2013*a*) or increased competition (Caron *et al.*, 2015).

(2) Thermal time and plant reproduction

All the climate change disturbances described above are ultimately a function of environmental temperature, and thus are suitable for thermal time modelling (Donohue et al., 2015). In a thermal time framework (also referred to as growing degree-day), physiological processes involve the accumulation of thermal time units towards the completion of a developmental phase. This phase can be flowering, attaining seed physiological maturity, or germination. Measuring the reproductive process in thermal time has the advantage of integrating time and temperature into a single physiological currency (Romo & Eddleman, 1995), and provides quantifiable units for the emerging framework of ecological energetics (Tomlinson et al., 2014). Five thermal time traits suffice to describe the thermal control of each reproductive step: the three cardinal temperatures or thermal thresholds (base, optimal, and ceiling) and the thermal time (sub- and supraoptimal) required to reach completion. These thermal time traits have two powerful applications. First, parameters calculated in a restricted set of treatments give experimental comprehensive estimations of reproduction in complex environments (Hardegree et al., 1999). This allows, for example, an exploration of plant reproduction in climate change scenarios (Orrù et al., 2012). Second, thermal time parameters are comparable in a standard way across seed individuals, populations and species (Trudgill, Squire & Thompson, 2000). As such, thermal time traits provide the broadest base to conduct comparative studies and metaanalyses of germination (Arène et al., 2017), as well as to integrate reproduction into broader fields of modelling such as community assembly (Larson & Funk, 2016) or species distribution models (Bykova et al., 2012).

Individual models for almost all the individual steps of plant reproduction already exist. In agronomy, there is a long tradition of using growing degree-day models to predict crop phenology (Nesmith & Bridges, 1992) and yield (Ney & Turc, 1993). García-Huidobro *et al.* (1982) developed a thermal time model to describe the rate of germination as a function of temperature, and many authors have subsequently applied this concept to estimate the field emergence of weeds (Steinmaus, Prather & Holt, 2000). Later research extended the framework to physiologically dormant wild species, considering dormancy loss in thermal time terms (Pritchard, Tompsett & Manger, 1996). Gama-Arachchige *et al.* (2013) modelled periodical changes in coat permeability (i.e. physical dormancy). Most recently, Porceddu *et al.* (2017) made a first attempt to model embryo growth (i.e. morphological dormancy) by quantifying cardinal temperatures specific to embryo growth inside the seed. Postgermination seedling growth also has its own specific thermal time traits (Adam *et al.*, 2007).

All these approaches have in common the use of thermal time traits to quantify the temperature stimuli that the plant or seed is instantaneously experiencing. The main issue we now want to highlight is that thermal time traits are themselves dependent on thermal history, i.e. the temperatures experienced by the plant or seed – or by its ancestors – during previous life stages. In this way, plants have a 'thermal memory' that can keep track of long-term climatic trends (Trewavas, 2003). Therefore, an integrated model of plant reproduction needs to account for the interactive effects of present thermal environment and past thermal history. To contribute to this task, we conducted a meta-analysis of thermal memory in plant reproductive traits.

II. META-ANALYSIS METHODOLOGY

(1) Literature search

We conducted a meta-analysis of the thermal memory of plant reproductive traits following the principles outlined by Koricheva, Gurevitch & Mengersen (2013). The objective of the metaanalysis was to test whether there is a relationship between seed traits and thermal history, either directly or through other seed traits. To gather a comprehensive sample of the available evidence, we performed a literature search in the Thomson Reuters Web of Science[™]. We constructed a Boolean search string (see online Supporting information, Appendix S1) by grouping key words in three blocks: (1) climate change, (2) thermal time models, and (3) the relationship between the maternal thermal environment and seed production, dispersal, dormancy, germination, and seedling development. We chose not to review seed longevity in the soil, as this has been the subject of a recent comprehensive review (Long et al., 2015). In addition, because many works have studied the effect of post-dispersal temperature in regulating dormancy loss and germination, for these traits we restricted the search to those investigations that used a thermal time approach. Furthermore, we restricted the search to journal articles written in English and classified in a relevant subject area (listed in Appendix S1).

In April 2016, we ran a search whose steps are documented in a PRISMA chart provided as Appendix S2 (Moher *et al.*, 2009). The search returned 10,242 results. We did not add articles

known to us but not found by the search, because the objective was to obtain a representative and unbiased sample of current knowledge, rather than to collect every source. We performed four consecutive screenings of the results. In a first screening of titles and a second screening of abstracts, we removed non-relevant studies. The title screening reduced the list to 1,487 articles, and the abstract screening to 533. In a third screening, we extracted the following information from the abstract: (1) the reproductive trait or process studied; (2) the factor exerting an effect on this trait or process, which could be temperature or another trait; (3) whether the study reported an experimental manipulation of temperature or not; and (4) whether the temperature represented an instantaneous response to the current thermal environment, or a response to the previous thermal history experienced by the plant (i.e. thermal memory). We then removed articles not reporting experimental manipulation of temperature or not addressing the effect of thermal memory, reducing the list to 205 articles. We performed a fourth and final screening of the text of these articles to determine which had actual measurements of temperature and plant traits. In the fourth screening, we rejected 94 articles because they did not provide data of interest; and another 30 articles not reporting sufficient information to perform a weighted metaanalysis (i.e. the standard deviation or an alternative way of calculating it). We included data for 'wildtype' Arabidopsis thaliana laboratory populations but excluded mutants with seed functions supressed.

(2) Data extraction and calculation of effect sizes

Since the goal of our meta-analysis was to test the relationship between pairs of quantitative variables (e.g. maternal temperature and seed mass), the most appropriate effect size measure was Pearson's correlation coefficient, r (Rosenberg, Rothstein & Gurevitch, 2013). Experiments in our selection of references, however, presented two caveats to the application of r. Firstly, most of the experiments treated the experimental variable (e.g. temperature) as a categorical variable with two or a few experimental levels. Secondly, our study variables were not expected to have a linear relationship; but rather a unimodal one. As an example, the value of seed mass should peak at the optimal maternal temperature and decrease at sub-optimal and supraoptimal temperatures. If an article used three temperature levels representing the optimum, suboptimum, and supra-optimum; analysing the overall effect size could indicate no relationship; when in fact the data showed both a positive relationship (between the sub-optimum and the optimum) and a negative relationship (between the optimum and the supra-optimum).

Taking these caveats into consideration, we adopted the following procedure to extract effect sizes. When an article reported values for more than one species, population, or seed accession; we treated these values as separate experimental units nested within the reference. When an article treated the experimental variable as a factor with more than two levels, we only used the two treatments that produced the highest and the lowest value of the study variable (or the two extreme treatments in the case that the study variable showed no change). This assumed that the highest value was close to the optimum, and the lowest value represented a sub- or supra-optimal situation. While it is true that the true optimum might have been somewhere between the highest and lowest values, this should not affect the main outcome of the analysis, which aimed to detect a relationship rather than to define its cardinal values (i.e. the optimum, base and ceiling treatments) or its strength (i.e. the slope). In other words, what was relevant was whether the regression coefficient was different from zero, rather than its actual value. In practice, this approach means that the relationships might have been stronger than reported here.

For these two treatments we extracted the mean, standard deviation, and number of replicates; or calculated them from other parameters if absent. When the study variable was the germination proportion or percentage, we used the mean germination and the number of seeds to create 2×2 contingency tables. In a few cases, the reference treated the experimental variable as a quantitative variable and reported the full set of values in a figure or table; we then calculated *r* directly using the full data set. When any of the aforementioned measurements were provided graphically, we used WebPlotDigitizer v3.12 (Rohatgi, 2017) to extract the values. We compiled the extracted measures in a table (Appendix S3). We transformed the extracted values to effect sizes (Fisher's Z transformation of r) and weights (the inverse of Zr's variance) using the packages 'esc' (Lüdecke, 2017) and 'metafor' (Viechtbauer, 2010) of R (R Core Team, 2017) (see script in Appendix S4). The calculated effect sizes are provided in Appendix S5. When calculating the effect sizes, we considered the lower value of the experimental variable as the control, and the higher value as the treatment. Therefore, positive effect indicated a positive or sub-optimal sizes relationship, and negative effect sizes a negative or supra-optimal relationship. Since our main objective was to test for a relationship independently of the sign, we transformed all effect sizes to absolute values to pool them in the analyses; but retained the information on whether they were derived from a positive, negative, or neutral (no effect) relationship.

(3) Phylogenetic correlation matrices

The calculated effect sizes could not be regarded as independent, as they were from plant species linked by a common evolutionary history (Chamberlain et al., 2012; Lajeunesse, Rosenberg & Jennions, 2013). Therefore, before proceeding with the analyses, we constructed a phylogenetic correlation matrix. We extracted a phylogenetic tree including all the species in our analysis from the R20120829 tree of Phylomatic v3 (Webb & Donoghue, 2005). We adjusted branch lengths with the exponential node ages of Bell, Soltis & Soltis (2010), using the ages file provided by Gastauer & Meira-Neto (2016) and the 'bladj' function of Phylocom (Webb, Ackerly & Kembel, 2008). The tree is provided in Appendix S6. Finally, we created phylogenetic correlation matrices using 'geiger' (Harmon et al., 2008). We adjusted one such matrix to the specific list of species included in each of the models described below.

(4) Multi-variate meta-analysis models

The nature of our data set, encompassing different plant species from different ecosystems, as well as laboratory and field experiments, made it highly unlikely that the effect sizes shared a common true mean. Furthermore, the data set had two sources of non-independence. Firstly, as discussed above, phylogenetic relatedness. Secondly, the fact that some studies provided more than one experimental unit (species, populations, or seed accessions). We therefore chose a multi-level or hierarchical random model to analyse our effect sizes (Koricheva et al., 2013). We fitted separate models to each relationship between pairs of variables, as there was relatively little overlap between studies and effect sizes. Each model included a random factor for reference, a nested random factor for experimental unit within reference, and a correlated error structure represented by the phylogenetic correlation matrix. We fitted the models using the rma.mv function of 'metafor' (see script in Appendix S7 and tabulated model outputs in Appendix S8).

(5) Heterogeneity in effect sizes

To estimate the heterogeneity or inconsistency in effect sizes (Senior *et al.*, 2016), we calculated two statistics: Q, which tests whether there is a significant amount of variance among studies; and I^2 , which indicates which percentage of the total variance can be attributed to the different random terms in the model (Nakagawa & Santos, 2012; Nakagawa *et al.*, 2017). Considering the low number of references for some of our relationships, and wanting to avoid over-complicated models, we decided against including explicit moderators in the models to explore the causes of heterogeneity. Instead, we fitted separate models and calculated

effect sizes for subgroups of the data representing different groupings. The first grouping considered the nature of the relationship (i.e. the original sign of the effect size), which could be positive/sub-optimal, neutral, or negative/supra-optimal. The second grouping considered the type of study species as reported in the reference: (*i*) wild, including weeds and exotics; (*ii*) domesticated, including crops and plantation trees; and (*iii*) Arabidopsis thaliana 'wildtype' laboratory populations, which represented a sizable portion of the data set and can be argued to be either wild or domesticated. The third grouping considered the experimental setting, either the laboratory (including greenhouses and experimental agricultural fields) or natural field sites.

(6) Publication bias

To test for publication bias or small-study effect we used the random-effects version of Egger's regression test (Rothstein *et al.*, 2005). Because the standard error of our effect size *r* is a function of the effect size itself, we used as a precision measure the inverse of the sample size (Peters *et al.*, 2006). Thus, we refitted the multi-variate meta-analysis models using the inverse of the sample size as a moderator. A relationship between the effect size and the precision measure could be interpreted as an indication of publication bias.

(7) Sensitivity analysis

Sensitivity analysis for multi-variate meta-analysis models is an active field of research. Following the procedure of recent investigations in plant sciences (Habeck & Schultz, 2015), we conducted ours by refitting the models after removing influential and outlier effect sizes. We defined influential effect sizes as those with hat values (i.e. diagonal elements of the hat matrix) greater than two times the average hat value; and outlier effect sizes as those with standardised residual values exceeding 3. We also tested the sensitivity of the analysis to the phylogenetic component by fitting alternative models with a tree calibrated with log-normal rather than exponential ages; as well as models without a phylogenetic correlation structure.

III. META-ANALYSIS OF THE THERMAL MEMORY OF PLANT REPRODUCTIVE TRAITS

(1) General meta-analysis diagnostics

Our literature search resulted in a set of 81 articles representing 73 taxa from 26 plant families. These articles are marked with asterisks in the reference list. From these articles, we were able to extract 433 effect-size records for the following 11 relationships:

maternal temperature with seed mass, seed number, germination proportion, time to 50% germination, and seedling growth; seed mass with germination proportion, time to 50% germination, and seedling growth; dormancy loss treatment with base germination temperature, base germination water potential, and thermal time for 50% germination. We conducted a meta-analysis (i.e. a quantitative synthesis) of these 11 relationships. In addition, during the literature search we found reports of other relationships or aspects of thermal memory that are worth considering as part of a qualitative synthesis (Appendix S2), even though we were unable to extract the data to include them in the meta-analysis. In the following sections, we combine the quantitative and qualitative syntheses in a narrative account of the successive processes of plant reproduction by seed, including the analysis of effect sizes and significant references for other processes that could not be analysed. First, we present the assessment of heterogeneity, publication bias and sensitivity.

Effect sizes had a generally high level of heterogeneity (Figs 1-3), as may be expected for a data set representing species across the diversity of seed plants. The only cases in which heterogeneity (as measured by Q) was not significant were the relationships of maternal temperature with seedling growth, and dormancy status with base water potential. These two cases were small data subsets dominated by a few species. For the most part, the major source of heterogeneity as estimated by I^2 was at the reference level. Phylogenetic relatedness was the largest contributor to heterogeneity in the relationships between seed mass and germination proportion and time, as well as dormancy and base germination temperature. Given the heterogeneity inherent in our data set, we believe that these heterogeneity values are only an indication of the patterns in the specific set of effect sizes that we gathered, rather than an indication of any general trend. Consistent with expectations, there was a consistent pattern of larger effect-size averages in laboratory vs. field experiments (Figs 1–3), although the confidence intervals largely overlapped. The categories (wild, domesticated species and Arabidopsis) tended to have the same effect sizes. Only one relationship (between maternal temperature and germination proportion) showed a stronger effect in the sub-optimal range of temperatures, coinciding with a large difference in the number of records (117 sub-optimal versus 8 supra-optimal).

The regression test for publication bias was negative in all cases (P > 0.10), indicating no relationship between outcome and precision. The sensitivity analysis identified 44 out of 433 records as influential, and one record as an outlier. The models fitted without these records, with an alternative phylogenetic tree, or with no tree (Appendix S9) did not deviate much from the conclusions of the full models, other than by an increased effect size or reduced confidence interval. Such differences are discussed in the specific sections below.

(2) Reproductive phenology

Several studies suggested that the phenology of reproduction (flowering and seed development) has thermal memory. For example, previous exposure to cold temperatures can enhance or be a requisite for flowering (Nesmith & Bridges, 1992). Flowering can be responsive to temperatures experienced as early as the seed maturation stage (Rubio de Casas *et al.*, 2012), or even by previous generations (Clapham, Willcott & Fedders, 2000). Furthermore, flowering time can be affected by seed mass (Elwell *et al.*, 2011). Such limited examples, however, meant that there was insufficient data to run a meta-analysis.

(3) Seed yield: mass and number

Thirty references from 30 species recorded the mass of seeds that had matured at different temperatures (Fig. 1A) with an even representation of records across temperature range, species category and experimental setting. The overall effect size was significantly different from zero and had a mean 68% correlation, indicating a strong thermal memory for seed mass. Similarly, 23 references from 22 species measured seed number in response to maternal temperature, finding an overall correlation of 68% (Fig. 1B). In this case there was a preponderance of supra-optimal records coming from investigations of extraordinary warming on crop yield. Both the higher contribution of phylogeny to the heterogeneity of mass, and the high inconsistency of number across studies, support the general assumption that seed number is a more plastic trait than seed mass (Baker, 2004). Within a given species, both components of yield generally have neutral or positive correlations, as both depend positively on plant size and resource availability (Venable, 1992).

Although the data made it impossible to dissect the different stages of maturation, individual references showed that the temperature dependence of yield can be specific to very precise stages, e.g. to tissue differentiation (early development events) but less so to filling and maturation (late events) (Wagner & Mitterhofer, 1998). Furthermore, thermal memory can last several years in masting tree populations (Kelly *et al.*, 2013), and be observed generations after in seed mass (Elwell *et al.*, 2011) and number (Bloedner *et al.*, 2007).

(4) Seed dispersal

Dispersability depends on diaspore traits such as terminal velocity, floatability and seed shape (Jacobs & Lesmeister, 2012) which depend strongly on yield traits. In particular, lighter seeds disperse longer distances (Ghamari *et al.*, 2010), while more seeds increase the chances of long-distance dispersal events (Higgins & Richardson, 1999). While we did not find studies testing the direct effect of maternal temperature on dispersal distance, it is clear that there is an indirect effect of thermal memory through seed yield.

(5) Morphological dormancy and embryo growth

Morphological dormancy is a trait in which the seed cannot germinate at the time of dispersal because the embryo is undifferentiated or underdeveloped. Seed maternal temperatures could affect morphological dormancy by determining: (a) the degree of embryo differentiation/development at dispersal; and/or (b) the rate of embryo growth in response to postdispersal temperatures. Evidence of this is, however, scarce. Mondoni et al. (2008) compared morphological dormancy between mountain and lowland populations of the temperate woodland forb Anemone nemorosa. Embryo size at dispersal was similar in both populations, but embryo growth was faster in the mountain population. Unfortunately, it was not possible to conclude whether this difference in growth rate was due to different seed development temperatures (i.e. thermal memory), or to genetic differences among sites.

(6) Physical dormancy and coat permeability

Physical dormancy is a block to seed imbibition (and consequently, to germination) imposed by an impermeable seed coat. Coat impermeability to water results from one or more layers of palisade cells in the testa, and in certain cases by specialised structures that regulate water uptake such as the hilum. It is relevant to consider whether seed development temperature affect could the development of these structures, thus determining the degree of physical dormancy at dispersal. There is evidence for variation in physical dormancy among individuals of the same population (Liyanage & Ooi, 2015), but as yet no proof that this variation is influenced by seed maternal temperatures.

(7) Primary physiological seed dormancy and germination traits

The level of physiological dormancy is difficult to measure *per se*, with the majority of studies quantifying it as a lack of germination. Thus, physiological dormancy and germinability become two faces of the same phenomenon, which can be measured in a variety of traits that include the germination proportion, germination rate or speed, cardinal temperatures, and thermal time. Predispersal thermal memory could affect these traits in two ways. First, it could determine their values at the time of dispersal (i.e. primary dormancy level). Second, it could determine their sensitivity to dormancy loss. It is important to note that this second case is a different situation from the effect of postdispersal temperatures driving dormancy loss and reimposition (see Section III.8).

The germination proportion (or final germination percentage) is by far the most widely used measure of seed dormancy and germinability. Twenty-seven studies of 24 taxa investigated the germination proportion of seeds from different maternal temperatures (Fig. 1C). Out of 156 records, a sizable proportion arose from laboratory Arabidopsis experiments. Most of the records indicated a positive relationship, and the average effect size was higher for sub-optimal temperature increases. It seems well established that warmer maternal environments result in higher germination (Fenner, 1991), but it is reasonable to assume that extraordinary warming could impair viability. A characteristic of germination proportion data sets was a high number of neutral or no-effect records, coming from experiments in which germination was 0 or 100% at both temperatures. Individual studies also suggested that seeds lose dormancy faster in response to warmer maternal environments (Hoyle et al., 2008a), although there were not enough data for meta-analysis. Six references of seven species analysed the effect of maternal temperatures on germination time (Fig. 1D), again with a majority of laboratory Arabidopsis trials. Although we found a significant overall effect size (P = 0.012), the confidence intervals were wider than for other relationships, and significance was lost when influential effect sizes were excluded (Appendix S9, Fig. S7D). We found no articles that measured the effect of maternal temperature on hydrothermal traits.

We were able to conduct a meta-analysis on the effect of seed mass on the germination proportion (23 studies, 24 species) and time (8 studies, 9 species) (Fig. 2A, B). These relationships had a high contribution of phylogeny to heterogeneity, perhaps as a result of constraints on seed mass. The effect for germination proportion was the same as for the relationship between temperature and proportion (38%). For germination time, the number of records was low, heterogeneity high, and the effect non-significantly different from zero (although it was marginally significant when using an alternative phylogenetic tree, and highly significant when considering no phylogenetic relatedness or excluding influential and outlier effect sizes,

Appendix S9, Figs S2B, S5B, S8B). We found a few reports of seed mass affecting hydrothermal time traits, with heavier seeds having lower base temperatures for germination (Wang, Bai & Tanino, 2004) and lower base water potentials (Brunel *et al.*, 2009).

(8) Post-dispersal seed dormancy cycles

Once dispersed, seeds undergo cyclic changes in their dormancy levels. Depending on their postdispersal thermal history, these can decrease (dormancy loss) or increase (secondary dormancy). Our meta-analysis showed that these changes are in fact due to alteration of the germination hydrothermal parameters (Fig. 3A, B): accumulation of time in dormancy loss conditions changes the germination base temperature (8 references, 7 species), base water potential (10 references, 8 species) and required thermal time for 50% germination (5 references, 8 species). Negative effect sizes here represent a widening of the conditions that will make the seed germinate: as dormancy is released, seeds can germinate at lower temperatures and water potentials, and require less accumulation of thermal time. Conversely, positive relationships indicate that secondary dormancy operates by increasing the base temperature or the required thermal time. In fact, individual studies showed that not only long-term stratification or after-ripening function in this way. Similar changes in hydrothermal traits are elicited by other factors such as fluctuating temperatures (Fernández-Pascual, Seal & Pritchard, 2015) and light (Hu et al., 2013).

(9) Seedling growth

We found three references (four species) dealing with the effect of maternal temperatures on seedling growth. These references provided five records for a meta-analysis, which indicated a marginally significant effect (Fig. 1E). The effect of seed mass on seedling growth was the topic of four references from four species, with a stronger overall effect size (Fig. 2C).

IV. SYNTHESIS

(1) The first hub of thermal memory: seed yield

There is an extensive literature on the thermal memory of seed mass and number, and relating these two traits to a wide range of other reproductive traits (Fig. 4). It is beyond doubt that a warming climate will affect seed yield, and the key question is whether this warming will occur within the suboptimal temperature range or push species into the supra-optimal range. A specific case of concern is

determinate crops, for which it is widely expected that warming – in the absence of other limiting factors - will accelerate all reproductive phases and thus reduce yield (Wheeler et al., 2000). This reduction is expected to be greater for seed number, since this is a more plastic trait than seed mass, especially in crops that have been subject to artificial selection (Sadras, 2007). In non-determinate plants, growth depends on the length of the season with suitable temperatures, and thus sub-optimal warming could improve yields. A key goal for future research will thus be to measure the yield thermal thresholds for a variety of ecologically significant species. As we have seen, another knowledge gap to fill is to dissect the role of different stages of thermal history, including previous generations and specific stages of flowering and seed development (Wagner & Mitterhofer, 1998). Another important step will be to integrate the thermal memory of seed yield into projections of species dispersal and future distributions. Yield improvements can reduce the dispersability of individual seeds (by increasing their mass and terminal velocity) while increasing the chances of an individual dispersing a small fraction of its progeny to long distances (by increasing its seed number, and likely seedling vigour). The overall outcome of these two seemingly opposite trends is not clear. The effect of mass on germination and seedling growth also needs clarification. For example, we found reports of heavier seeds having lower germination, but it is unclear whether this indicates that they are less viable or have deeper dormancy. At least in some cases, heavier and moredeveloped seeds seem to have deeper dormancy (Wang et al., 2010). It is also unclear if there is a causal relationship between mass and germination, or an indirect correlation because both traits are influenced by maternal temperature. Overall, our meta-analysis reflects the importance of seed mass as a central hub for studies of the thermal memory of plant regeneration. In community ecology, articles considering physical seed traits, in particular seed mass, have increased fourfold in the last ten years (Jiménez-Alfaro et al., 2016). This preponderance of reports on seed mass presumably reflects the ease of measurement of this trait as opposed to seed physiological thresholds and thermal times.

(2) The second hub of thermal memory: seed dormancy

The study of germination as a response to postdispersal temperatures has reached maturity within the thermal time approach, and thermal time traits are accumulating for an ever-increasing number of species (Seal *et al.*, 2017). Nonetheless, the study of pre-dispersal temperatures and the thermal memory of germination lags behind. As we have seen, most experiments investigating maternal temperature have measured germination as the final proportion of germinated seeds, a trait which provides little information on the relationships between temperature and reproduction. In a way, pre- and post-dispersal studies are speaking different languages: the former describing proportions and the latter thermal-time. This is a huge handicap to the incorporation of thermal memory into post-dispersal emergence models. It is important for future research on the effect of maternal temperature to adopt a hydrothermal approach, to allow us to develop integrated pre-dispersal/post-dispersal modelling of seed dormancy and germination.

We show one such model in Fig. 5. At the most basic level, germination thermal traits respond to the immediate post-dispersal thermal environment and drive progress towards germination. A first complication involves post-dispersal dormancy cycling, which produces changes in the hydrothermal thresholds and thermal time required for seed germination. During dormancy loss, the thresholds widen and the thermal time required is reduced, hence seeds become 'easier to germinate'; the opposite occurs during the imposition of secondary dormancy (Jones, Ellis & Gosling, 1997). In terms of thermal time, the rate of dormancy change is a function of exposure to post-dispersal temperatures (Pritchard et al., 1996). The rate of dormancy loss is proportional to the difference between the actual temperature and a thermal threshold for dormancy change (Ceccato, Daniel Bertero & Batlla, 2011), and thus can be incorporated into the model by considering a separate set of dormancy-change thermal traits.

Further development of the model allows the integration of pre-dispersal maternal temperatures, which can affect: (1) the initial level of dormancy at dispersal; and (2) the rate of dormancy loss. This second aspect particularly requires further investigation, as little is known about the relationship between maternal temperature and the thermal thresholds for dormancy loss. One likely hypothesis is that these thresholds (for example, the ceiling temperature below which dormancy loss can occur) are dependent on the maternal temperatures experienced during seed development. Much remains to be clarified regarding the thermal memory of physiological dormancy, and very few data are available for species with physically and morphologically dormant seeds.

(3) Implications for modelling: an example

The thermal memory of plant reproduction has profound ecological implications, especially in climate change scenarios, where it has the potential to facilitate acclimatisation to new environments. For example, one concern is that future winters may not be cold enough to break physiological dormancy (Orrù *et al.*, 2012). Warmer climates could also

result in more shallow physiological dormancy (Huang *et al.*, 2018). To illustrate the importance of this point, we provide an example in which future seed germination is projected under six different models that differ in their assumptions regarding thermal memory (Fig. 6). To simplify matters, in these models we show changes produced by thermal memory only as changes in the thermal time required for 50% germination (θ_{50}), although in a real scenario both θ_{50} and the cardinal temperatures could change.

In the simplest model with non-dormant seeds (model ND0 in Fig. 6), dispersed seeds accumulate thermal time (red triangles) until they reach θ_{50} (green lines), at which point they germinate. In a warmer future (model ND1), thermal time accumulates faster, leading to earlier seed germination. We can extend this model by including the effect of pre-dispersal thermal memory on seed mass (see Fig. 4) (model ND2). If we assume that a future increase in temperature will be in the sub-optimal range, this would lead to heavier seeds. If we also assume that heavier seeds germinate faster (Fig. 4), we can expect θ_{50} to be reduced, and thus earlier germination than in model ND1.

A more realistic simulation for many plant species should incorporate the role of physiological seed dormancy. In our first example (model D0), dormancy is represented by a decrease in θ_{50} during exposure to cold winter temperatures (blue bars). In practice, this means that, after chilling, seeds need to accumulate less thermal time to germinate. If a warmer future reduces the length of the overwintering season, dormancy loss could be compromised (model D1). After a shortened winter, θ_{50} would remain too high, potentially preventing the seed from germinating. In the final model (model D2), we integrate the effect of thermal memory on seed mass and dormancy (see Fig. 5). In this case, a warmer future would produce seeds that start with a lower θ_{50} . Even if the amount of dormancy loss they receive decreases because of a shortened winter, it might be sufficient to reduce θ_{50} to a level that allows adequate timing of germination. The key difference between D1 and D2 is within the seed: in both scenarios, the external environment provides the same signals for dormancy loss and germination. Through thermal memory, D2 seeds can adjust their 'expectations' to the new climate. D1 seeds can not, and thus fail to germinate.

In conclusion, thermal memory should allow seeds to adjust their germination phenology to climatic changes. In this example we have seen how increasing environmental temperatures can reduce the amount of cold stratification that a seed *receives*, but also the amount it *needs to receive* to break dormancy. This feedback will likely ensure some successful reproduction even if warming goes beyond the current thermal thresholds. In any event, the effect of temperatures experienced during maternal can influence future germination traits, and climate change simulations should take this into account.

(4) Future priorities

One role for biology in these times of global change is to provide general rules that predict the behaviour of complex ecosystems. A key tool in this pursuit is trait-based models, which provide a predictive relationships framework for the between biodiversity and the environment (Funk et al., 2017). For plant reproduction by seed, incorporating quantitative physiological traits into multi-species studies remains a difficult task, hindered by the lack of reliable trait databases and comparable methodologies for wild species (Jiménez-Alfaro et al., 2016). In stark contrast with other functional traits, there is no standard template for physiological thermal-time studies. A widely followed handbook on the measurement of plant functional traits (Pérez-Harguindeguy et al., 2013) deals only with seed mass and shape, and does not mention germination as a trait. Thus, seed ecology remains fundamentally disconnected from comparative and functional ecology. A modelling approach such as that described herein, applied to a global range of species characteristic of well-defined vegetation types, could provide a basis for a research agenda on global plant reproduction.

Ultimately, understanding plant reproduction by seed means being able to answer several questions. When are seeds produced? How many and how heavy are these seeds? How far do they disperse in space, but also in time through seed dormancy? When do they germinate? How many achieve successful seedling establishment? None of these questions have binary answers, all are a matter of degree. Their quantitative answers depend on physiological seed traits: the cardinal temperatures and thermal times of the different reproductive processes. These traits integrate the climatic inputs of the environment to produce reproductive outputs in the form of timing and success. It is crucial to understand these traits not as fixed numbers, but as having their own thermal memory, sometimes extending over several generations. This complex system - in which seed thermal-time traits are not static but in permanent change as a function of thermal history – ensures phenological integration of plant reproduction. We suggest that such a system will offer a degree of resilience and acclimatisation to climate change, until certain tipping points are passed, and reproduction becomes unsuccessful. Identifying these resilience thresholds remains a chief task for plant biologists and global change researchers. We believe that meeting this challenge will move us towards a more quantitative framework

that sees changes in plant life not in absolute terms of positives and negatives, but as the acceleration or deceleration of physiological rates.

V. CONCLUSIONS

(1) Plant reproduction by seed is a complex process that integrates past and present climatic inputs to achieve successful establishment outputs. The successive steps of seed production, dispersal, dormancy, and germination respond to instantaneous thermal cues; but they also have a thermal memory (*via* phenotypic plasticity) that incorporates information from past thermal history.

(2) Seed yield is the first hub of thermal memory. Models of future dispersal and distribution should consider how the future climate will affect seed mass and number, as these two traits determine the dispersability of individuals and populations, as well as the vigour of their seedlings.

(3) Seed dormancy is the second hub of thermal memory, playing a key role in the acclimatisation of establishment timing. Research must link predispersal and post-dispersal thermal histories by using a common thermal-time framework. Physical and morphological dormancy warrant particular efforts.

(4) This integrated system makes plant reproduction from seed relatively resilient to climatic changes, as long as certain tipping points are not passed. The thermal time approach offers a valuable methodological framework to identify such tipping points. Future progress will depend on the adoption of a standardised and simplified methodology for the calculation of thermal time traits, and the application of this methodology to large numbers of ecologically relevant species in a range of habitats.

VI. ACKNOWLEDGEMENTS

We would like to thank several anonymous reviewers who contributed significant to improvements. Some drawings used in Figs 4-6 were downloaded from 'Pixabay - Stunning Free Images' (https://pixabay.com/) available under a CC0 Creative Commons Licence as free for commercial use with no attribution required. E.F.P. received financial support from the Government of Asturias and the FP7 – Marie Curie - COFUND programme of the European Commission (Grants 'Clarín' ACA14-19 and ACB17-19). H.W.P. acknowledges funding from the People Programme (Marie Curie Actions) of the European Union's Seventh Framework Programme FP7/2007-2013/ under REA grant agreement no. 607785 for the NASSTEC project. The Royal Botanic Gardens, Kew, receive grant-in-aid from Defra.

VII. REFERENCES

An asterisk (*) indicates that the reference was included in the meta-analysis.

- * ADAM, N.R., DIERIG, D.A., COFFELT, T.A., WINTERMEYER, M.J., MACKEY, B.E. & WALL, G.W. (2007). Cardinal temperatures for germination and early growth of two *Lesquerella species*. *Industrial Crops and Products* **25**, 24–33.
- * ALATALO, J.M. & TOTLAND, O. (1997). Response to simulated climatic change in an alpine and subarctic pollen-risk strategist, *Silene acaulis*. *Global Change Biology* 3, 74–79.
- * ALVARADO, V. & BRADFORD, K.J. (2005). Hydrothermal time analysis of seed dormancy in true (botanical) potato seeds. *Seed Science Research* **15**, 77–88.
- ARÈNE, F., AFFRE, L., DOXA, A. & SAATKAMP, A. (2017). Temperature but not moisture response of germination shows phylogenetic constraints while both interact with seed mass and lifespan. *Seed Science Research* 27, 110– 120.
- * AUGE, G.A., BLAIR, L.K., BURGHARDT, L.T., COUGHLAN, J., EDWARDS, B., LEVERETT, L.D. & DONOHUE, K. (2015). Secondary dormancy dynamics depends on primary dormancy status in *Arabidopsis thaliana*. *Seed Science Research* 25, 230–246.
- AUGE, G.A., LEVERETT, L.D., EDWARDS, B.R. & DONOHUE, K. (2017). Adjusting phenotypes via within- and across-generational plasticity. *New Phytologist* **216**, 343–349.
- * BAIR, N.B., MEYER, S.E. & ALLEN, P.S. (2006). A hydrothermal after-ripening time model for seed dormancy loss in *Bromus tectorum* L. *Seed Science Research* **16**, 17–28.
- BAKER, J.T. (2004). Yield responses of Southern US rice cultivars to CO2 and temperature. *Agricultural and Forest Meteorology* **122**, 129–137.
- * BALOCH, H.A., DITOMMASO, A. & WATSON, A.K. (2001). Intrapopulation variation in *Abutilon theophrasti* seed mass and its relationship to seed germinability. *Seed Science Research* 11, 335–343.
- * BATLLA, D. & BENECH-ARNOLD, R.L. (2003). A quantitative analysis of dormancy loss dynamics in *Polygonum aviculare* L. seeds: Development of a thermal time model based on changes in seed population thermal parameters. *Seed Science Research* **13**, 55–68.
- * BATLLA, D. & BENECH-ARNOLD, R.L. (2004). A

predictive model for dormancy loss in *Polygonum aviculare* L. seeds based on changes in population hydrotime parameters. *Seed Science Research* **14**, 277–286.

- * BAUER, M.C., MEYER, S.E. & ALLEN, P.S. (1998). A simulation model to predict seed dormancy loss in the field for *Bromus tectorum* L. *Journal of Experimental Botany* **49**, 1235– 1244.
- BELL, C.D., SOLTIS, D.E. & SOLTIS, P.S. (2010). The age and diversification of the angiosperms rerevisited. *American Journal of Botany* 97, 1296–1303.
- * BERNAREGGI, G., CARBOGNANI, M., PETRAGLIA, A. & MONDONI, A. (2015). Climate warming could increase seed longevity of alpine snowbed plants. *Alpine Botany* **125**, 69–78.
- BJORKMAN, A.D., ELMENDORF, S.C., BEAMISH, A.L., VELLEND, M. & HENRY, G.H.R. (2015). Contrasting effects of warming and increased snowfall on Arctic tundra plant phenology over the past two decades. *Global Change Biology* 21, 4651–4661.
- * BLÖDNER, C., GOEBEL, C., FEUSSNER, I., GATZ, C. & POLLE, A. (2007). Warm and cold parental reproductive environments affect seed properties, fitness, and cold responsiveness in *Arabidopsis thaliana* progenies. *Plant Cell and Environment* **30**, 165–175.
- * BOCHENEK, A., GOLASZEWSKI, J. & GIELWANOWSKA, I. (2010). Hydrotime model analysis of *Matricaria maritima* ssp. *inodora* seed dormancy. *Plant Species Biology* 25, 136–148.
- * BODDY, L.G., BRADFORD, K.J. & FISCHER, A.J. (2013). Stratification requirements for seed dormancy alleviation in a wetland weed. *Plos One* **8**.
- BRUNEL, S., TEULAT-MERAH, B., WAGNER, M.H., HUGUET, T., PROSPERI, J.M. & DUERR, C. (2009). Using a model-based framework for analysing genetic diversity during germination and heterotrophic growth of *Medicago truncatula*. *Annals of Botany* **103**, 1103–1117.
- * BRUNEL-MUGUET, S., D'HOOGHE, P., BATAILLE, M.-P., LARRE, C., KIM, T.-H., TROUVERIE, J., AVICE, J.-C., ETIENNE, P. & DUERR, C. (2015). Heat stress during seed filling interferes with sulfur restriction on grain composition and seed germination in oilseed rape (*Brassica* napus L.). Frontiers in Plant Science 6, 1-12.
- BYKOVA, O., CHUINE, I., MORIN, X. & HIGGINS, S.I. (2012). Temperature dependence of the reproduction niche and its relevance for plant species distributions. *Journal of Biogeography*

39, 2191–2200.

- CARON, M.M., DE FRENNE, P., BRUNET, J., CHABRERIE, O., COUSINS, S.A.O., DECOCQ, G., DIEKMANN, M., GRAAE, B.J., HEINKEN, T., KOLB, A., LENOIR, J., NAAF, T., PLUE, J., SELVI, F., WULF, M., *ET AL*. (2015). Divergent regeneration responses of two closely related tree species to direct abiotic and indirect biotic effects of climate change. *Forest Ecology and Management* **342**, 21–29.
- * CASTOLDI, E. & MOLINA, J.A. (2014). Effect of seed mass and number of cotyledons on seed germination after heat treatment in *Pinus* sylvestris L. var. *iberica* Svob. *Forest Systems* 23, 483–489.
- CECCATO, D. V, DANIEL BERTERO, H. & BATLLA, D. (2011). Environmental control of dormancy in quinoa (*Chenopodium quinoa*) seeds: two potential genetic resources for pre-harvest sprouting tolerance. *Seed Science Research* **21**, 133–141.
- CHAMBERLAIN, S.A., HOVICK, S.M., DIBBLE, C.J., RASMUSSEN, N.L., VAN ALLEN, B.G., MAITNER, B.S., AHERN, J.R., BELL-DERESKE, L.P., ROY, C.L., MEZA-LOPEZ, M., CARRILLO, J., SIEMANN, E., LAJEUNESSE, M.J. & WHITNEY, K.D. (2012). Does phylogeny matter? Assessing the impact of phylogenetic information in ecological meta-analysis. *Ecology Letters* **15**, 627–636.
- * CHANTRE, G.R., BATLLA, D., SABBATINI, M.R. & ORIOLI, G. (2009). Germination parameterization and development of an afterripening thermal-time model for primary dormancy release of *Lithospermum arvense* seeds. *Annals of Botany* **103**, 1291–1301.
- * CHANTRE, G.R., SABBATINI, M.R. & ORIOLI, G.A. (2010). An after-ripening thermal-time model for *Lithospermum arvense* seeds based on changes in population hydrotime parameters. *Weed Research* **50**, 218–227.
- CHEESMAN, A.W. & WINTER, K. (2013). Elevated night-time temperatures increase growth in seedlings of two tropical pioneer tree species. *New Phytologist* **197**, 1185–1192.
- * CHIANG, G.C.K., BARTSCH, M., BARUA, D., NAKABAYASHI, K., DEBIEU, M., KRONHOLM, I., KOORNNEEF, M., SOPPE, W.J.J., DONOHUE, K. & DE MEAUX, J. (2011). DOG1 expression is predicted by the seed-maturation environment and contributes to geographical variation in germination in *Arabidopsis thaliana*. *Molecular Ecology* **20**, 3336–3349.
- * CHRISTENSEN, M., MEYER, S.E. & ALLEN, P.S. (1996). A hydrothermal time model of seed after-ripening in *Bromus tectorum* L. Seed

Science Research 6, 155–163.

- CLAPHAM, W.M., WILLCOTT, J.B. & FEDDERS, J.M. (2000). Effects of seed maturation temperature on seed yield characteristics and subsequent generations of lupin. *Crop Science* **40**, 1313– 1317.
- COCHRANE, A., DAWS, M.I. & HAY, F.R. (2011). Seed-based approach for identifying flora at risk from climate warming. *Austral Ecology* **36**, 923–935.
- COCHRANE, A., YATES, C.J., HOYLE, G.L. & NICOTRA, A.B. (2015). Will amongpopulation variation in seed traits improve the chance of species persistence under climate change? *Global Ecology and Biogeography* **24**, 12–24.
- * DAWSON, T.E. & EHLERINGER, J.R. (1991). Ecological correlates of seed mass variation in *Phoradendron juniperinum*, a xylem-tapping mistletoe. *Oecologia* **85**, 332–342.
- * DECHAINE, J.M., GARDNER, G. & WEINIG, C. (2009). Phytochromes differentially regulate seed germination responses to light quality and temperature cues during seed maturation. *Plant Cell and Environment* **32**, 1297–1309.
- * DE FRENNE, P., BRUNET, J., SHEVTSOVA, A., KOLB, A., GRAAE, B.J., CHABRERIE, O., COUSINS, S.A., DECOCQ, G., DE SCHRIJVER, A., DIEKMANN, M., GRUWEZ, R., HEINKEN, T., HERMY, M., NILSSON, C., STANTON, S., *ET AL.* (2011). Temperature effects on forest herbs assessed by warming and transplant experiments along a latitudinal gradient. *Global Change Biology* **17**, 3240–3253.
- DE FRENNE, P., GRAAE, B.J., BRUNET, J., SHEVTSOVA, A., DE SCHRIJVER, A., CHABRERIE, O., COUSINS, S.A.O., DECOCQ, G., DIEKMANN, M., HERMY, M., HEINKEN, T., KOLB, A., NILSSON, C., STANTON, S. & VERHEYEN, K. (2012). The response of forest plant regeneration to temperature variation along a latitudinal gradient. *Annals of Botany* 109, 1037–1046.
- DEL CACHO, M., ESTIARTE, M., PEÑUELAS, J. & LLORET, F. (2013*a*). Inter-annual variability of seed rain and seedling establishment of two woody Mediterranean species under field-induced drought and warming. *Population Ecology* **55**, 277–289.
- * DEL CACHO, M., PEÑUELAS, J. & LLORET, F. (2013b). Reproductive output in Mediterranean shrubs under climate change experimentally induced by drought and warming. *Perspectives in Plant Ecology Evolution and Systematics* **15**, 319–327.

- * DELGADO, J.A., SERRANO, J.M., LOPEZ, F. & ACOSTA, F.J. (2008). Seed size and seed germination in the Mediterranean fire-prone shrub *Cistus ladanifer*. *Plant Ecology* **197**, 269–276.
- DONOHUE, K., BURGHARDT, L.T., RUNCIE, D., BRADFORD, K.J. & SCHMITT, J. (2015). Applying developmental threshold models to evolutionary ecology. *Trends in Ecology & Evolution* **30**, 66–77.
- * DONOHUE, K., HESCHEL, M.S., CHIANG, G.C.K., BUTLER, C.M. & BARUA, D. (2007). Phytochrome mediates germination responses to multiple seasonal cues. *Plant Cell and Environment* **30**, 202–212.
- ELWELL, A.L., GRONWALL, D.S., MILLER, N.D., SPALDING, E.P. & BROOKS, T.L.D. (2011). Separating parental environment from seed size effects on next generation growth and development in *Arabidopsis*. *Plant Cell and Environment* **34**, 291–301.
- ENGLER, R., RANDIN, C.F., VITTOZ, P., CZAKA, T., BENISTON, M., ZIMMERMANN, N.E. & GUISAN, A. (2009). Predicting future distributions of mountain plants under climate change: does dispersal capacity matter? *Ecography* 32, 34–45.
- FENNER, M. (1991). The effects of the parent environment on seed germinability. *Seed Science Research* **1**, 75–84.
- FERNÁNDEZ-PASCUAL, E., JIMÉNEZ-ALFARO, B., CAUJAPÉ-CASTELLS, J., JAÉN-MOLINA, R. & DÍAZ, T.E. (2013). A local dormancy cline is related to the seed maturation environment, population genetic composition and climate. *Annals of Botany* **112**, 937–945.
- FERNÁNDEZ-PASCUAL, E., SEAL, C.E. & PRITCHARD, H.W. (2015). Simulating the germination response to diurnally alternating temperatures under climate change scenarios: comparative studies on *Carex diandra* seeds. *Annals of Botany* **115**, 201–209.
- * FIGUEROA, R., HERMS, D.A., CARDINA, J. & DOOHAN, D. (2010). Maternal Environment Effects on Common Groundsel (*Senecio* vulgaris) Seed Dormancy. Weed Science 58, 160–166.
- FUNK, J.L., LARSON, J.E., AMES, G.M., BUTTERFIELD, B.J., CAVENDER-BARES, J., FIRN, J., LAUGHLIN, D.C., SUTTON-GRIER, A.E., WILLIAMS, L. & WRIGHT, J. (2017). Revisiting the Holy Grail: using plant functional traits to understand ecological processes. *Biological Reviews* 92, 1156–1173.
- GAMA-ARACHCHIGE, N.S., BASKIN, J.M., GENEVE,

R.L. & BASKIN, C.C. (2013). Quantitative analysis of the thermal requirements for stepwise physical dormancy-break in seeds of the winter annual *Geranium carolinianum* (Geraniaceae). *Annals of Botany* **111**, 849–858.

- * GAO, S., WANG, J., ZHANG, Z., DONG, G. & GUO, J. (2012). Seed production, mass, germinability, and subsequent seedling growth responses to parental warming environment in *Leymus chinensis. Crop & Pasture Science* 63, 87–94.
- GARCIA-HUIDOBRO, J., MONTEITH, J.L. & SQUIRE, G.R. (1982). Time, temperature and germination of pearl-millet (*Pennisetum typhoides* S. & H.) .1. Constant temperature. *Journal of Experimental Botany* **33**, 288–296.
- GASTAUER, M. & MEIRA-NETO, J.A.A. (2016). An enhanced calibration of a recently released megatree for the analysis of phylogenetic diversity. *Brazilian Journal of Biology* **76**, 619–628.
- GHAMARI, S., BORGHEI, A.M., RABBANI, H., KHAZAEI, J. & BASATI, F. (2010). Modeling the terminal velocity of agricultural seeds with artificial neural networks. *African Journal of Agricultural Research* 5, 389–398.
- * GIANINETTI, A. & COHN, M.A. (2007). Seed dormancy in red rice. XII: Population-based analysis of dry-afterripening with a hydrotime model. *Seed Science Research* 17, 253–271.
- * GOLOMBEK, S.D., SULTANA, A. & JOHANSEN, C. (2001). Effect of separate pod and root zone temperatures on yield and seed composition of three Spanish cultivars of groundnut (*Arachis hypogaea* L). *Journal of the Science of Food and Agriculture* **81**, 1326–1333.
- * GRASS, L. & BURRIS, J.S. (1995). Effect of heat stress during seed development and maturation on wheat (*Triticum durum*) seed quality .1. Seed germination and seedling vigor. *Canadian Journal of Plant Science* **75**, 821– 829.
- * GUTTERMAN, Y. (2001). Phenotypic germination plasticity related to caryopsis size in *Schismus arabicus*. *Seed Science Research* **11**, 173–178.
- HABECK, C.W. & SCHULTZ, A.K. (2015). Community-level impacts of white-tailed deer on understorey plants in North American forests: a meta-analysis. *AoB Plants* **7**, plv119.
- * HARDEGREE, S.P. & VAN VACTOR, S.S. (2000). Germination and emergence of primed grass seeds under field and simulated-field temperature regimes. *Annals of Botany* **85**, 379–390.

- HARDEGREE, S.P., VAN VACTOR, S.S., PIERSON, F.B. & PALMQUIST, D.E. (1999). Predicting variable-temperature response of non-dormant seeds from constant-temperature germination data. *Journal of Range Management* 52, 83– 91.
- HARMON, L.J., WEIR, J.T., BROCK, C.D., GLOR, R.E. & CHALLENGER, W. (2008). GEIGER: investigating evolutionary radiations. *Bioinformatics* 24, 129–131.
- * HE, H., VIDIGAL, D. DE S., SNOEK, L.B., SCHNABEL, S., NIJVEEN, H., HILHORST, H. & BENTSINK, L. (2014). Interaction between parental environment and genotype affects plant and seed performance in *Arabidopsis*. *Journal of Experimental Botany* **65**, 6603– 6615.
- HIGGINS, S.I. & RICHARDSON, D.M. (1999). Predicting plant migration rates in a changing world: the role of long-distance dispersal. *The American Naturalist* **153**, 464–475.
- * HOYLE, G.L., DAWS, M.I., STEADMAN, K.J. & ADKINS, S.W. (2008*a*). Pre- and post-harvest influences on physiological dormancy alleviation of an Australian Asteraceae species: *Actinobole uliginosum* (A. Gray) H. Eichler. *Seed Science Research* **18**, 191–199.
- * HOYLE, G.L., STEADMAN, K.J., DAWS, M.I. & ADKINS, S.I.W. (2008b). Pre- and post-harvest influences on seed dormancy status of an Australian Goodeniaceae species, *Goodenia fascicularis*. *Annals of Botany* **102**, 93–101.
- HU, X.W., ZHOU, Z.Q., LI, T.S., WU, Y.P. & WANG, Y.R. (2013). Environmental factors controlling seed germination and seedling recruitment of *Stipa bungeana* on the Loess Plateau of northwestern China. *Ecological Research* 28, 801–809.
- * HUANG, Z., FOOTITT, H.O., FOOTITT, S. & FINCH-SAVAGE, W.E. (2015). Seed dormancy is a dynamic state: variable responses to preand post-shedding environmental signals in seeds of contrasting *Arabidopsis* ecotypes. *Seed Science Research* **25**, 159–169.
- * HUANG, Z., FOOTITT, S. & FINCH-SAVAGE, W.E. (2014). The effect of temperature on reproduction in the summer and winter annual *Arabidopsis thaliana* ecotypes Bur and Cvi. *Annals of Botany* **113**, 921–929.
- HUANG, Z., FOOTITT, S., TANG, A. & FINCH-SAVAGE, W.E. (2018). Predicted global warming scenarios impact on the mother plant to alter seed dormancy and germination behaviour in *Arabidopsis*. *Plant, Cell & Environment* **41**, 187–197.

- HUANG, Z., LIU, S., BRADFORD, K.J., HUXMAN, T.E. & VENABLE, D.L. (2016). The contribution of germination functional traits to population dynamics of a desert plant community. *Ecology* 97, 250–261.
- HUELBER, K., BARDY, K. & DULINGER, S. (2011). Effects of snowmelt timing and competition on the performance of alpine snowbed plants. *Perspectives in Plant Ecology Evolution and Systematics* **13**, 15–26.
- * HUME, L. (1994). Maternal environment effects on plant-growth and germination of 2 strains of *Thlaspi arvense* L. *International Journal of Plant Sciences* **155**, 180–186.
- JACOBS, B.S. & LESMEISTER, S.A. (2012). Maternal environmental effects on fitness, fruit morphology and ballistic seed dispersal distance in an annual forb. *Functional Ecology* 26, 588–597.
- * JENSEN, M. & ERIKSEN, E.N. (2001). Development of primary dormancy in seeds of *Prunus avium* during maturation. *Seed Science and Technology* **29**, 307–320.
- JIMÉNEZ-ALFARO, B., SILVEIRA, F.A.O., FIDELIS, A., POSCHLOD, P. & COMMANDER, L.E. (2016). Seed germination traits can contribute better to plant community ecology. *Journal of Vegetation Science* 27, 637–645.
- JONES, S.K., ELLIS, R.H. & GOSLING, P.G. (1997). Loss and induction of conditional dormancy in seeds of *Sitka spruce* maintained moist at different temperatures. *Seed Science Research* 7, 351–358.
- * JUMRANI, K. & BHATIA, V.S. (2014). Impact of elevated temperatures on growth and yield of chickpea (*Cicer arietinum* L.). *Field Crops Research* **164**, 90–97.
- * KAUR, R., BAINS, T.S., BINDUMADHAVA, H. & NAYYAR, H. (2015). Responses of mungbean (*Vigna radiata* L.) genotypes to heat stress: Effects on reproductive biology, leaf function and yield traits. *Scientia Horticulturae* **197**, 527–541.
- KELLY, D., GELDENHUIS, A., JAMES, A., HOLLAND, E.P., PLANK, M.J., BROCKIE, R.E., COWAN, P.E., HARPER, G.A., LEE, W.G., MAITLAND, M.J., MARK, A.F., MILLS, J.A., WILSON, P.R. & BYROM, A.E. (2013). Of mast and mean: differential-temperature cue makes mast seeding insensitive to climate change. *Ecology Letters* 16, 90–98.
- * KLADY, R.A., HENRY, G.H.R. & LEMAY, V. (2011). Changes in high arctic tundra plant reproduction in response to long-term experimental warming. *Global Change*

Biology **17**, 1611–1624.

- KORICHEVA, J., GUREVITCH, J. & MENGERSEN, K.L. (2013). *Handbook of Meta-Analysis in Ecology and Evolution*. Princeton University Press.
- * KOZAREWA, I., CANTLIFFE, D.J., NAGATA, R.T. & STOFFELLA, P.J. (2006). High maturation temperature of lettuce seeds during development increased ethylene production and germination at elevated temperatures. *Journal of the American Society for Horticultural Science* **131**, 564–570.
- KUPARINEN, A., KATUL, G., NATHAN, R. & SCHURR, F.M. (2009). Increases in air temperature can promote wind-driven dispersal and spread of plants. *Proceedings of the Royal Society B-Biological Sciences* 276, 3081–3087.
- * LACEY, E.P. (1996). Parental effects in *Plantago lanceolata* L .1. A growth chamber experiment to examine pre- and postzygotic temperature effects. *Evolution* **50**, 865–878.
- LAJEUNESSE, M.J., ROSENBERG, M.S. & JENNIONS, M.D. (2013). Phylogenetic Nonindependence and Meta-analysis. In *Handbook of Metaanalysis in Ecology and Evolution* pp. 284– 299. Princeton University Press, Princeton, NJ.
- * LARSEN, S.U. & ANDREASEN, C. (2004). Light and heavy turfgrass seeds differ in germination percentage and mean germination thermal time. *Crop Science* **44**, 1710–1720.
- LARSON, J.E. & FUNK, J.L. (2016). Regeneration: an overlooked aspect of trait-based plant community assembly models. *Journal of Ecology* **104**, 1284–1298.
- LI, D.-Z. & PRITCHARD, H.W. (2009). The science and economics of *ex situ* plant conservation. *Trends in plant science* **14**, 614–621.
- * LIMBACH, W.E. & CALL, C.A. (1996). Germination response of Russian wildrye to variations in seed mass at fluctuating temperatures. *Journal of Range Management* 49, 346–349.
- LIYANAGE, G.S. & OOI, M.K.J. (2015). Intrapopulation level variation in thresholds for physical dormancy-breaking temperature. *Annals of Botany* **116**, 123–131.
- LONG, R.L., GORECKI, M.J., RENTON, M., SCOTT, J.K., COLVILLE, L., GOGGIN, D.E., COMMANDER, L.E., WESTCOTT, D.A., CHERRY, H. & FINCH-SAVAGE, W.E. (2015). The ecophysiology of seed persistence: a mechanistic view of the journey to germination or demise. *Biological Reviews* **90**, 31–59.

- LÜDECKE, D. (2017). esc: Effect Size Computation for Meta Analysis. R package version 0.4.0.
- * MACGREGOR, D.R., KENDALL, S.L., FLORANCE, H., FEDI, F., MOORE, K., PASZKIEWICZ, K., SMIRNOFF, N. & PENFIELD, S. (2015). Seed production temperature regulation of primary dormancy occurs through control of seed coat phenylpropanoid metabolism. *New Phytologist* **205**, 642–652.
- * MADAN, P., JAGADISH, S.V.K., CRAUFURD, P.Q., FITZGERALD, M., LAFARGE, T. & WHEELER, T.R. (2012). Effect of elevated CO2 and high temperature on seed-set and grain quality of rice. *Journal of Experimental Botany* 63, 3843–3852.
- * MALCOLM, P.J., HOLFORD, P., MCGLASSON, W.B. & NEWMAN, S. (2003). Temperature and seed weight affect the germination of peach rootstock seeds and the growth of rootstock seedlings. *Scientia Horticulturae* **98**, 247–256.
- MERRITT, D.J. & DIXON, K.W. (2011). Restoration seed banks—a matter of scale. *Science* **332**, 424–425.
- * MEYER, S.E., DEBAENE-GILL, S.B. & ALLEN, P.S. (2000). Using hydrothermal time concepts to model seed germination response to temperature, dormancy loss, and priming effects in *Elymus elymoides*. *Seed Science Research* **10**, 213–223.
- * MIDMORE, E.K., MCCARTAN, S.A., JINKS, R.L. & CAHALAN, C.M. (2015). Using thermal time models to predict germination of five provenances of silver birch (*Betula pendula* Roth) in southern England. *Silva Fennica* **49**.
- MILBAU, A., GRAAE, B.J., SHEVTSOVA, A. & NIJS, I. (2009). Effects of a warmer climate on seed germination in the subarctic. *Annals of Botany* 104, 287–296.
- MOHER, D., LIBERATI, A., TETZLAFF, J., ALTMAN, D.G. & GROUP, T.P. (2009). Preferred Reporting Items for Systematic Reviews and Meta-Analyses: The PRISMA Statement. *PLoS Medicine* 6, e1000097.
- * MOLAU, U. & SHAVER, G.R. (1997). Controls on seed production and seed germinability in *Eriophorum vaginatum*. *Global Change Biology* **3**, 80–88.
- MONDONI, A., PEDRINI, S., BERNAREGGI, G., ROSSI, G., ABELI, T., PROBERT, R.J., GHITTI, M., BONOMI, C. & ORSENIGO, S. (2015). Climate warming could increase recruitment success in glacier foreland plants. *Annals of Botany* **116**, mcv101.
- MONDONI, A., PROBERT, R., ROSSI, G., HAY, F. & BONOMI, C. (2008). Habitat-correlated seed

germination behaviour in populations of wood anemone (*Anemone nemorosa* L.) from northern Italy. *Seed Science Research* **18**, 213–222.

- * MU, J., PENG, Y. & NIU, K. (2013). Divergent seed production responses of white and blue flowers of *Gentiana leucomelaena* (Gentianaceae) to warming and watering. *Plant Ecology & Diversity* **6**, 495–501.
- * MURPHEY, M., KOVACH, K., ELNACASH, T., HE, H., BENTSINK, L. & DONOHUE, K. (2015). DOG1-imposed dormancy mediates germination responses to temperature cues. *Environmental and Experimental Botany* **112**, 33–43.
- MUTKE, S., GORDO, J. & GIL, L. (2005). Variability of Mediterranean Stone pine cone production: Yield loss as response to climate change. *Agricultural and Forest Meteorology* **132**, 263–272.
- NAKAGAWA, S., NOBLE, D.W.A., SENIOR, A.M. & LAGISZ, M. (2017). Meta-evaluation of metaanalysis: ten appraisal questions for biologists. *BMC Biology* **15**, 18.
- NAKAGAWA, S. & SANTOS, E.S.A. (2012). Methodological issues and advances in biological meta-analysis. *Evolutionary Ecology* **26**, 1253–1274.
- NESMITH, D.S. & BRIDGES, D.C. (1992). Modeling chilling influence on cumulative flowering - a case-study using tifblue rabbiteye blueberry. *Journal of the American Society for Horticultural Science* **117**, 698–702.
- NEY, B. & TURC, O. (1993). Heat-unit-based description of the reproductive development of pea. *Crop Science* **33**, 510–514.
- NICOTRA, A.B., ATKIN, O.K., BONSER, S.P., DAVIDSON, A.M., FINNEGAN, E.J., MATHESIUS, U., POOT, P., PURUGGANAN, M.D., RICHARDS, C.L. & VALLADARES, F. (2010). Plant phenotypic plasticity in a changing climate. *Trends in Plant Science* **15**, 684–692.
- OOI, M.K.J., AULD, T.D. & DENHAM, A.J. (2009). Climate change and bet-hedging: interactions between increased soil temperatures and seed bank persistence. *Global Change Biology* **15**, 2375–2386.
- ORRÙ, M., MATTANA, E., PRITCHARD, H.W. & BACCHETTA, G. (2012). Thermal thresholds as predictors of seed dormancy release and germination timing: altitude-related risks from climate warming for the wild grapevine *Vitis vinifera* subsp. *sylvestris. Annals of Botany* **110**, 1651–1660.

- PÉREZ-HARGUINDEGUY, N., DÍAZ, S., GARNIER, É., LAVOREL, S., POORTER, H., JAUREGUIBERRY, P., BRET-HARTE, M., CORNWELL, W.K., CRAINE, J. & GURVICH, D. (2013). New handbook for standardised measurement of plant functional traits worldwide. *Australian Journal of Botany* 61, 167–234.
- PETERS, J.L., SUTTON, A.J., JONES, D.R., ABRAMS, K.R. & RUSHTON, L. (2006). Comparison of Two Methods to Detect Publication Bias in Meta-analysis. JAMA 2952956.
- * PORCEDDU, M., MATTANA, E., PRITCHARD, H.W. & BACCHETTA, G. (2013). Thermal niche for in situ seed germination by Mediterranean mountain streams: model prediction and validation for *Rhamnus persicifolia* seeds. *Annals of Botany* **112**, 1887–1897.
- PORCEDDU, M., MATTANA, E., PRITCHARD, H.W. & BACCHETTA, G. (2017). Dissecting seed dormancy and germination in *Aquilegia barbaricina*, through thermal kinetics of embryo growth. *Plant Biology* **19**, 983–993.
- * PRASAD, P.V. V, BOOTE, K.J., ALLEN, L.H. & THOMAS, J.M.G. (2002). Effects of elevated temperature and carbon dioxide on seed-set and yield of kidney bean (*Phaseolus vulgaris* L.). *Global Change Biology* 8, 710–721.
- * PRASAD, P.V. V, DJANAGUIRAMAN, M., PERUMAL, R. & CIAMPITTI, I.A. (2015). Impact of high temperature stress on floret fertility and individual grain weight of grain sorghum: sensitive stages and thresholds for temperature and duration. *Frontiers in Plant Science* **6**, 1– 11.
- * PRASAD, P.V.V., BOOTE, K.J. & ALLEN JR., L.H. (2006). Adverse high temperature effects on pollen viability, seed-set, seed yield and harvest index of grain-sorghum *Sorghum bicolor* (L.) Moench are more severe at elevated carbon dioxide due to higher tissue temperatures. *Agricultural and Forest Meteorology* 139, 237–251.
- * PRITCHARD, H.W., STEADMAN, K.J., NASH, J. V & JONES, C. (1999). Kinetics of dormancy release and the high temperature germination response in *Aesculus hippocastanum* seeds. *Journal of Experimental Botany* **50**, 1507–1514.
- PRITCHARD, H.W., TOMPSETT, P.B. & MANGER, K.R. (1996). Development of a thermal time model for the quantification of dormancy loss in *Aesculus hippocastanum* seeds. *Seed Science Research* **6**, 127–135.
- * QADERI, M.M. & CAVERS, P.B. (2000). Variation in germination response within Scotch thistle, *Onopordum acanthium* L., populations

matured under greenhouse and field conditions. *Ecoscience* **7**, 57–65.

- * QADERI, M.M., CAVERS, P.B. & BERNARDS, M.A. (2003). Pre- and post-dispersal factors regulate germination patterns and structural characteristics of Scotch thistle (*Onopordum acanthium*) cypselas. *New Phytologist* **159**, 263–278.
- * QADERI, M.M., CAVERS, P.B., HAMILL, A.S., DOWNS, M.P. & BERNARDS, M.A. (2006). Maturation temperature regulates germinability and chemical constituents of Scotch thistle (*Onopordum acanthium*) cypselas. *Canadian Journal of Botany-Revue Canadienne De Botanique* **84**, 28–38.
- R CORE TEAM (2017). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. Https://www.r-project.org/.
- * RENZI, J.P., CHANTRE, G.R. & CANTAMUTTO, M.A. (2014). Development of a thermal-time model for combinational dormancy release of hairy vetch (*Vicia villosa* ssp. *villosa*). Crop & Pasture Science **65**, 470–478.
- RICHARDSON, S.J., ALLEN, R.B., WHITEHEAD, D., CARSWELL, F.E., RUSCOE, W.A. & PLATT, K.H. (2005). Climate and net carbon availability determine temporal patterns of seed production by *Nothofagus. Ecology* 86, 972–981.
- ROHATGI, A. (2017). WebPlotDigitizer. Austin, Texas. *WebPlotDigitizer*. Http://arohatgi.info/WebPlotDigitizer.
- ROMO, J.T. & EDDLEMAN, L.E. (1995). Use of degree-days in multiple-temperature experiments. *Journal of Range Management* 48, 410–416.
- * RONDANINI, D., SAVIN, R. & HALL, A.J. (2003). Dynamics of fruit growth and oil quality of sunflower (*Helianthus annuus* L.) exposed to brief intervals of high temperature during grain filling. *Field Crops Research* 83, 79–90.
- ROSENBERG, M.S., ROTHSTEIN, H.R. & GUREVITCH, J. (2013). Effect Sizes: Conventional Choices and Calculations. In *Handbook of Metaanalysis in Ecology and Evolution* pp. 61–71. Princeton University Press, Princeton, NJ.
- ROTHSTEIN, H., SUTTON, A.J., BORENSTEIN, M. & WILEY INTERSCIENCE (ONLINE SERVICE) (2005). Publication bias in meta-analysis: prevention, assessment and adjustments. Wiley.
- RUBIO DE CASAS, R., KOVACH, K., DITTMAR, E., BARUA, D., BARCO, B. & DONOHUE, K. (2012). Seed after-ripening and dormancy

determine adult life history independently of germination timing. *New Phytologist* **194**, 868–879.

- * RUIZ-VERA, U.M., SIEBERS, M.H., DRAG, D.W., ORT, D.R. & BERNACCHI, C.J. (2015). Canopy warming caused photosynthetic acclimation and reduced seed yield in maize grown at ambient and elevated CO2. *Global Change Biology* 21, 4237–4249.
- SADRAS, V.O. (2007). Evolutionary aspects of the trade-off between seed size and number in crops. *Field Crops Research* **100**, 125–138.
- * SANDVIK, S.M. & EIDE, W. (2009). Costs of reproduction in circumpolar *Parnassia palustris* L. in light of global warming. *Plant Ecology* **205**, 1–11.
- * SATO, S., KAMIYAMA, M., IWATA, T., MAKITA, N., FURUKAWA, H. & IKEDA, H. (2006). Moderate increase of mean daily temperature adversely affects fruit set of *Lycopersicon esculentum* by disrupting specific physiological processes in male reproductive development. *Annals of Botany* 97, 731–738.
- * SCHMUTHS, H., BACHMANN, K., WEBER, W.E., HORRES, R. & HOFFMANN, M.H. (2006). Effects of preconditioning and temperature during germination of 73 natural accessions of *Arabidopsis thaliana*. *Annals of Botany* **97**, 623–634.
- SEAL, C.E., DAWS, M.I., FLORES, J., ORTEGA-BAES, P., GALÍNDEZ, G., LEÓN-LOBOS, P., SANDOVAL, A., CERONI STUVA, A., RAMÍREZ BULLÓN, N., DÁVILA-ARANDA, P., ORDOÑEZ-SALANUEVA, C.A., YÁÑEZ-ESPINOSA, L., ULIAN, T., AMOSSO, C., ZUBANI, L., *ET AL.* (2017). Thermal buffering capacity of the germination phenotype across the environmental envelope of the Cactaceae. *Global Change Biology* 23, 5309–5317.
- SENIOR, A.M., GRUEBER, C.E., KAMIYA, T., LAGISZ, M., O'DWYER, K., SANTOS, E.S.A. & NAKAGAWA, S. (2016). Heterogeneity in ecological and evolutionary meta-analyses: its magnitude and implications. *Ecology* 97, 3293–3299. Wiley-Blackwell.
- * SHARIF-ZADEH, F. & MURDOCH, A.J. (2000). The effects of different maturation conditions on seed dormancy and germination of *Cenchrus ciliaris. Seed Science Research* **10**, 447–457.
- * SINGH, V. Y, NGUYEN, C.T., VAN OOSTEROM, E.J., CHAPMAN, S.C., JORDAN, D.R. & HAMMER, G.L. (2015). *Sorghum* genotypes differ in high temperature responses for seed set. *Field Crops Research* **171**, 32–40.
- * STEADMAN, K.J., ELLERY, A.J., CHAPMAN, R.,

MOORE, A. & TURNER, N.C. (2004). Maturation temperature and rainfall influence seed dormancy characteristics of annual ryegrass (*Lolium rigidum*). *Australian Journal* of Agricultural Research **55**, 1047–1057.

- * STEADMAN, K.J. & PRITCHARD, H.W. (2004). Germination of <u>Aesculus hippocastanum</u> seeds following cold-induced dormancy loss can be described in relation to a temperaturedependent reduction in base temperature (T_b) and thermal time. *New Phytologist* **161**, 415– 425.
- STEINMAUS, S.J., PRATHER, T.S. & HOLT, J.S. (2000). Estimation of base temperatures for nine weed species. *Journal of Experimental Botany* **51**, 275–286.
- * STENSTROM, A. & JONSDOTTIR, I.S. (2004). Effects of simulated climate change on phenology and life history traits in *Carex bigelowii*. *Nordic Journal of Botany* **24**, 355–371.
- STEVENS, N., SEAL, C.E., ARCHIBALD, S. & BOND, W. (2014). Increasing temperatures can improve seedling establishment in aridadapted savanna trees. *Oecologia* 175, 1029– 1040.
- * SUNG, Y., CANTLIFFE, D.J. & NAGATA, R.T. (1998). Seed developmental temperature regulation of thermotolerance in lettuce. *Journal of the American Society for Horticultural Science* **123**, 700–705.
- * SWAIN, A.J., HUGHES, Z.S., COOK, S.K. & MOSS, S.R. (2006). Quantifying the dormancy of *Alopecurus myosuroides* seeds produced by plants exposed to different soil moisture and temperature regimes. *Weed Research* 46, 470– 479.
- * TANVEER, A., TASNEEM, M., KHALIQ, A., JAVAID, M.M. & CHAUDHRY, M.N. (2013). Influence of seed size and ecological factors on the germination and emergence of field bindweed (*Convolvulus arvensis*). *Planta Daninha* **31**, 39–51.
- TOMLINSON, S., ARNALL, S.G., MUNN, A., BRADSHAW, S.D., MALONEY, S.K., DIXON, K.W. & DIDHAM, R.K. (2014). Applications and implications of ecological energetics. *Trends in Ecology & Evolution* **29**, 280–290.
- TREWAVAS, A. (2003). Aspects of plant intelligence. Annals of Botany **92**, 1–20.
- TRUDGILL, D.L., SQUIRE, G.R. & THOMPSON, K. (2000). A thermal time basis for comparing the germination requirements of some British herbaceous plants. *New Phytologist* **145**, 107–114.
- VENABLE, D.L. (1992). Size-number trade-offs and

the variation of seed size with plant resource status. *American Naturalist* **140**, 287–304.

- VIECHTBAUER, W. (2010). Conducting metaanalyses in R with the metafor package. *Journal of Statistical Software* **36**, 1–48.
- WAGNER, J. & MITTERHOFER, E. (1998). Phenology, seed development, and reproductive success of an alpine population of *Gentianella germanica* in climatically varying years. *Botanica Acta* 111, 159–166.
- WALCK, J.L., HIDAYATI, S.N., DIXON, K.W., THOMPSON, K. & POSCHLOD, P. (2011). Climate change and plant regeneration from seed. *Global Change Biology* **17**, 2145–2161.
- * WANG, A.B., TAN, D.Y., BASKIN, C.C. & BASKIN, J.M. (2010). Effect of seed position in spikelet on life history of *Eremopyrum distans* (Poaceae) from the cold desert of north-west China. *Annals of Botany* **106**, 95–105.
- WANG, R., BAI, Y. & TANINO, K. (2004). Effect of seed size and sub-zero imbibition-temperature on the thermal time model of winterfat (*Eurotia lanata* (Pursh) Moq.). *Environmental* and Experimental Botany **51**, 183–197.
- WEBB, C.O., ACKERLY, D.D. & KEMBEL, S.W. (2008). Phylocom: software for the analysis of phylogenetic community structure and trait evolution. *Bioinformatics* 24, 2098–2100.
- WEBB, C.O. & DONOGHUE, M.J. (2005). Phylomatic: tree assembly for applied phylogenetics. *Molecular Ecology Notes* 5, 181–183.
- * WEBBER, J., OTT, P., OWENS, J. & BINDER, W. (2005). Elevated temperature during reproductive development affects cone traits and progeny performance in *Picea glauca* x *engelmannii* complex. *Tree Physiology* **25**, 1219–1227.
- * WEI, Y., BAI, Y. & HENDERSON, D.C. (2009). Critical conditions for successful regeneration of an endangered annual plant, *Cryptantha minima*: A modeling approach. *Journal of Arid Environments* **73**, 872–875.
- WHEELER, T.R., CRAUFURD, P.Q., ELLIS, R.H., PORTER, J.R. & PRASAD, P.V. V (2000). Temperature variability and the yield of annual crops. Agriculture Ecosystems & Environment 82, 159–167.
- WOOKEY, P.A., PARSONS, A.N., WELKER, J.M., POTTER, J.A., CALLAGHAN, T. V, LEE, J.A. & PRESS, M.C. (1993). Comparative responses of phenology and reproductive development to simulated environmental-change in sub-arctic and high arctic plants. *Oikos* 67, 490–502.

- * ZHANG, J.H. & HAMILL, A.S. (1997). Seed weight, intraspecific competition, and plant performance in *Abutilon theophrasti*. *Canadian Journal of Botany-Revue Canadienne De Botanique* **75**, 1614–1620.
- * ZHANG, R., GALLAGHER, R.S. & SHEA, K. (2012). Maternal warming affects early life stages of an invasive thistle. *Plant Biology* **14**, 783–788.

VIII. SUPPORTING INFORMATION

Additional supporting information may be found in the online version of this article.

Appendix S1. Boolean search string used in the *Web* of *Science*.

Appendix S2. PRISMA chart showing the flow of information through the phases of the literature review.

Appendix S3. Data set with the values extracted from the references used in the meta-analysis. This data set can be used together with the R script in Appendix S4 to repeat the effect size calculations.

Appendix S4. R script used to calculate effect sizes from the data in Appendix S2.

Appendix S5. Data set with the calculated effect sizes for the studies in the meta-analysis, with information on the family, species accession number/strain, experimental setting, temperature ranges investigated and wild/domesticated status. This data set can be used together with the phylogenetic tree in Appendix S6 and the R script in Appendix S7 to repeat the meta-analysis model fitting.

Appendix S6. Newick phylogenetic tree used to fit the meta-analysis models.

Appendix S7. R script used to fit the meta-analysis models to the effect sizes in Appendix S5.

Appendix S8. The full output of the meta-analysis models.

Appendix S9. Results of the sensitivity analyses. Figures showing the results of models fitted with a different phylogenetic tree (Figs S1–S3), with no tree (Figs S4–S6), or by removing influential and outlier effect sizes (Figs S7–S9).



Fig. 1 Results of the meta-analyses for the effect of maternal temperature on several seed traits. The forest plots on the left show the estimated effect sizes (Pearson's correlation coefficient) with their 95% confidence intervals. The overall effect for the whole data set is represented by red diamonds. The partial effects according to different groupings of the data set (experimental setting, type of species and type of effect) are represented by circles of different colours. The number of review units (seed accessions within references) that contributed to the calculation of the effect size of each grouping is given to the right of the y axis. The tables on the right give the main parameters of each model: z is the Z-statistic, k is the number of review units in the model, I2 and Q are heterogeneity parameters. The total heterogeneity I2 can be calculated by adding the values for Phylogeny, Reference and Accession.



(A) Seed mass and germination proportion

Fig. 2. Results of the meta-analyses for the effect of seed mass on several seed traits. The forest plots on the left show the estimated effect sizes (Pearson's correlation coefficient) with their 95% confidence intervals. The overall effect for the whole data set is represented by red diamonds. The partial effects according to different groupings of the data set (experimental setting, type of species and type of effect) are represented by circles of different colours. The number of review units (seed accessions within references) that contributed to the calculation of the effect size of each grouping is given to the right of the y axis. The tables to the right give the main parameters of each model: z is the Z-statistic, k is the number of review units in the model, I2 and Q are heterogeneity parameters. The total heterogeneity I2 can be calculated adding the values for Phylogeny, Reference and Accession.



Fig. 3. Results of the meta-analyses for the effect of change in dormancy status on several seed traits. The forest plots on the left show the estimated effect sizes (Pearson's correlation coefficient) with their 95% confidence intervals. The overall effect for the whole data set is represented by red diamonds. The partial effects according to different groupings of the data set (experimental setting, type of species and type of effect) are represented by circles of different colours. The number of review units (seed accessions within references) that contributed to the calculation of the effect size of each grouping is given to the right of the y axis. The tables to the right give the main parameters of each model: z is the Z-statistic, k is the number of review units in the model, I2 and Q are heterogeneity parameters. The total heterogeneity I2 can be calculated adding the values for Phylogeny, Reference and Accession.

(A) Dormancy and base temperature



Fig. 4. Thermal memory of seed yield. Temperature increases in the sub-optimal range result in the production of more, heavier seeds; while the opposite occurs in the supra-optimal range. In turn, these two yield traits have a lasting effect on subsequent stages of plant reproduction. In the case of dispersal, increases of temperature in the sub-optimal range can decrease the dispersability of individual seeds, but increase the chance of some seeds dispersing over long distances. Sub-optimal range temperature increases also tend to favour faster seed germination and seedling establishment.



Fig. 5. Thermal memory of seed dormancy and germination. Post-dispersal temperatures (e.g. dormancy loss temperatures experienced in the soil seed bank) change the hydrothermal thresholds and the thermal time required for germination. Pre-dispersal temperatures (e.g. maternal temperatures experienced during seed maturation) can affect hydrothermal traits in two ways: (1) through an effect of seed mass on the hydrothermal traits; (2) through seed dormancy, as maternal temperature could (i) set the level of dormancy that the seeds have at dispersal; and (ii) set the sensitivity to dormancy loss/induction in response to the post-dispersal temperatures. In a practical scenario, seeds maturing in a warmer environment would have less dormancy at dispersal, and would lose this dormancy faster. As dormancy is released, seeds would become able to germinate at wider ranges of temperature and water potential, and would require less accumulation of thermal time before they germinate.



Fig. 6 Modelling of seed germination under climate change scenarios. Six models are presented that differ in their assumptions about seed dormancy and thermal memory. Field emergence of non-dormant seeds (ND) can be modelled using the cumulative sum of environmental temperatures (red areas) and the median thermal time for germination (θ 50, dashed green lines). The case of dormant seeds (D) can be considered by incorporating a chilling period (blue bars) that reduces θ 50. These models can take into account present temperatures (models ND0 and D0) and future climate change scenarios. They can ignore pre-dispersal thermal memory (models ND1 and D1) or take it into account (models ND2 and D2). In this simplification, dormancy loss and thermal memory are assumed to affect only the value of θ 50. Arrows highlight changes due to the predicted increase in environmental temperatures (e.g. from ND0 to ND1) or, within future scenarios, due to the inclusion of thermal memory (e.g. from ND1 to ND2). Grey areas and arrows represent previous conditions and directions of changes. See Section IV.3 for further discussion. The germinated seed indicates the moment of germination, in model D1 the seed fails to germinate due to insufficient dormancy loss.